

# *Propagation of hemp (Cannabis sativa)*

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**Cornell CALS**  
College of Agriculture and Life Sciences

Cornell University Hemp Webinar Series  
February 8, 2023







# ***LIHREC has specialists in all areas of horticulture:***

**Greenhouse & Floriculture  
Fruit: Viticulture  
Vegetables/potatoes  
Woody ornamentals/Landscape plants**

## **Cross Commodity Specialists:**

**Plant disease  
Weed science  
Entomology  
Plant Tissue Culture**

**We are  
one-of-a-kind  
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# • Plant Propagation



**Propagation media**



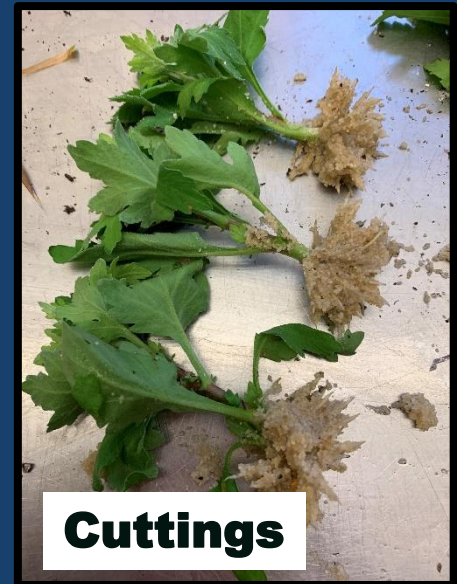
**Bud graft**



**Grafting**



**Air Layering**



**Cuttings**



# Plant Propagation

The multiplication of plants by both sexual and asexual means





# Hemp plants can be propagated by sexual and asexual means

## TODAY:

1. Seed propagation
2. Vegetative cutting propagation
3. Micropropagation



# HOW TO PROPAGATE PLANTS – THE BASICS

- Select **healthy specimens!**



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  - Retention of nutrient





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- **Light**
  - Daylength/Photoperiod
  - Intensity
  - Wavelength



# HOW TO PROPAGATE PLANTS – THE BASICS

- **Water management and humidity**
  - Intermittent mist?
  - Plastic covers?





# HOW TO PROPAGATE PLANTS – THE BASICS

- Water management and humidity
- Greenhouse Temperature
  - 73 - 77°F is ideal for germination and rooting of cuttings of most plants
    - For hemp: up to 80°F to 82°F



# HOW TO PROPAGATE PLANTS – THE BASICS

- Water management and humidity
- Greenhouse Temperature
- Cleanliness
  - Good sanitation



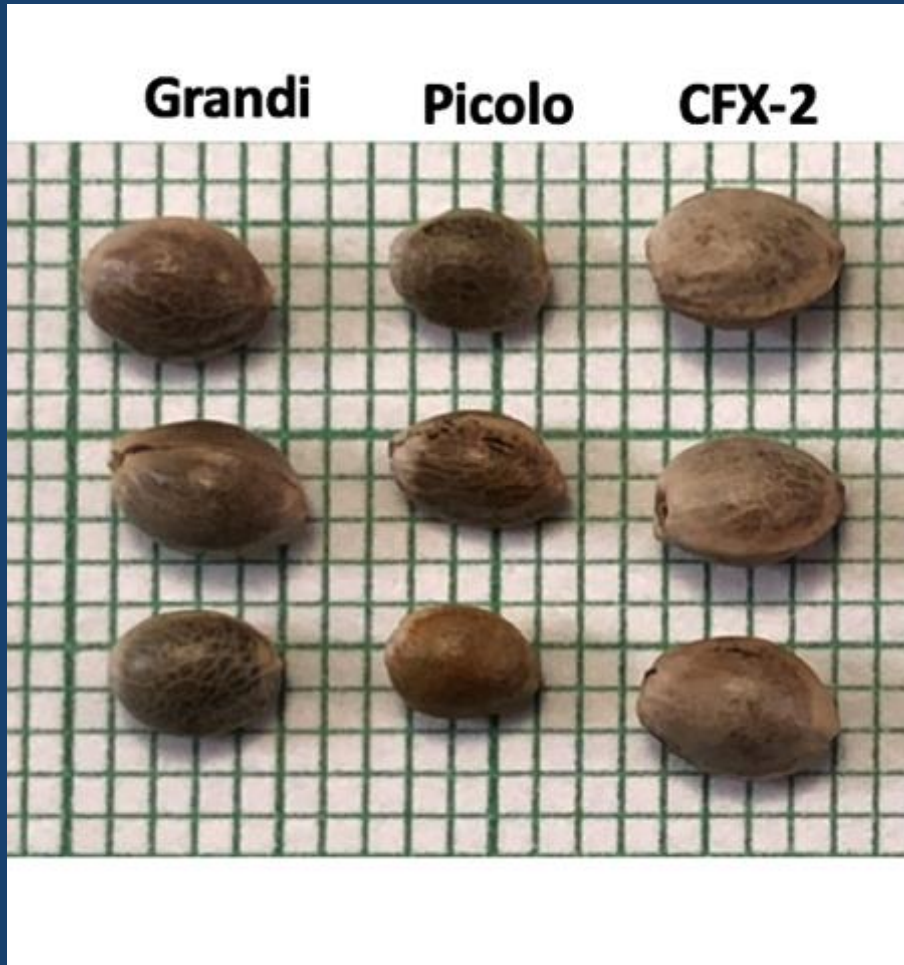


# Seed Propagation vs. Vegetative Propagation

- Hemp plants are **dioecious**
- Pistillate and staminate organs **occur on different plants.**
  - Pistillate plants are desirable
- Genetic recombination results in **segregation of traits & phenotypic diversity**
- Do you want clones?  
**Use vegetative cutting propagation**



# Seed Morphology



Dispersal unit is an achene - outer layer is a pericarp  
as in sunflower seed



# Seed Propagation Hemp

- Start with the correct, viable seed: use **GOOD GENETICS**
  - Feminized seeds for CBD hemp



# Seed Propagation Hemp

- Understand your **field soil or greenhouse media**:
  - well-draining, low weed pressure, and has plenty of nutrients will lay a healthy foundation
  - Soil tests for field soil





# Seed Propagation Hemp

- **Strategically plan** outdoor production to maximize the environment at your location
- Consider frosts, day length, and rain



# Seed Propagation Hemp

- Pre-soak seeds for 8-12 hours to improve germination (no longer than 24 hours)
  - Helpful, but not necessary
- Ideal seed germination temperature: Between 65°-80°F
- Plant seeds about 1-inch deep
- Seeds can germinate in light or dark conditions (Small 2016)
- Water thoroughly

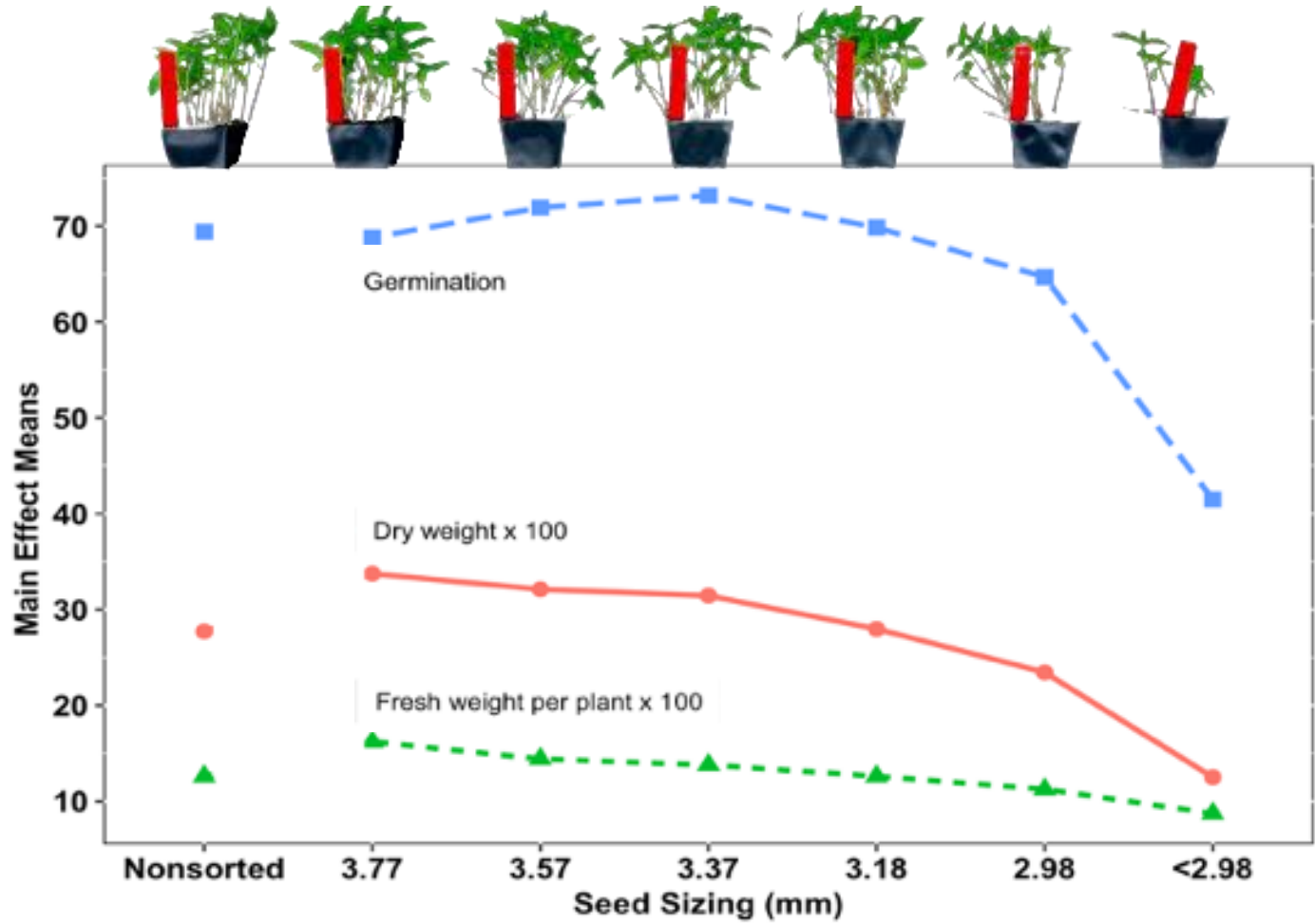




# Propagation from seed

Alan Taylor, Michael Loos,  
Masi Amirkhani, Hilary Mayton  
Cornell University, Geneva, NY

## SEED SIZE impacts seed quality



Credit Mike Quade 2022



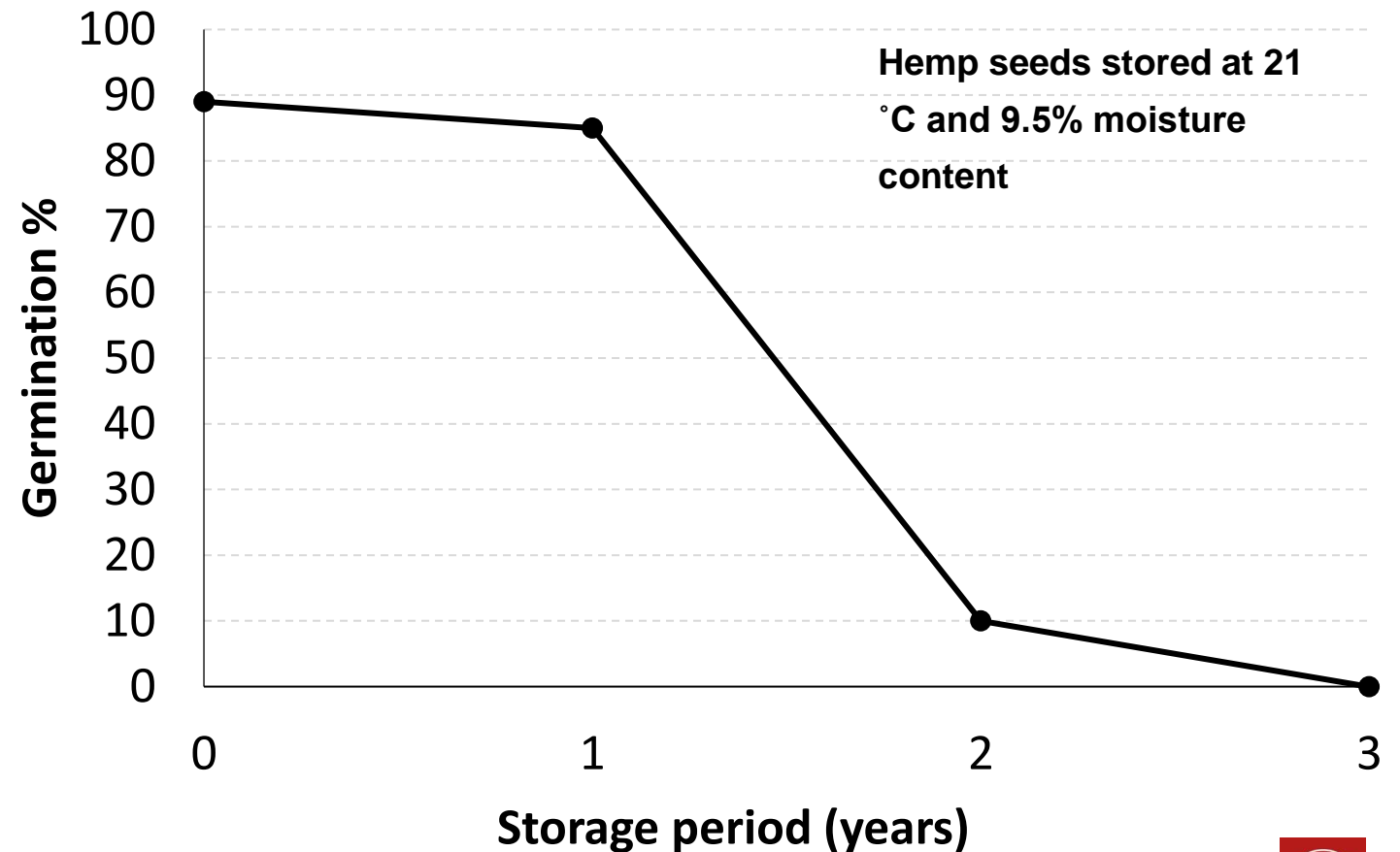
# Hemp Seed Storage

- Need to store seeds to maintain viability and vigor
- Hemp seed have short-medium longevity in relation to vegetable crop seeds

Alan Taylor, Michael Loos,  
Masi Amirkhani, Hilary Mayton  
Cornell University, Geneva, NY



Tetrazolium viability test. A, non-viable seeds, seed of hemp (Photo by S. Elias, Oregon State Univ).





# VEGETATIVE PROPAGATION\*

## WHY?

- A clone is an exact copy
  - to **maintain** their **genetic and phenotypic characteristics**



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- **Uniformity**





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- **Low cost**



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- Clones **get a head-start** compared to seedlings
- **Year-round production** in greenhouses



# What are STEM CUTTINGS?

- **Stem cuttings** = the most common approach to vegetative propagation hemp
- A cutting is a piece of plant that has been cut off from a parent plant and then **given the opportunity to make roots** of its own
- Grow into an **identical plant** as the original stock plant



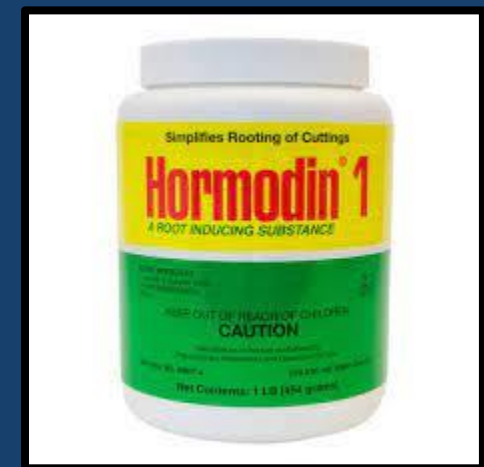


# What you will need:

- Sharp knife, clippers (scissors), or a razor
  - clean and disinfect all your tools
- Propagation medium
- Rooting hormone (1,000 ppm)
- Humidity domes or intermittent mist system (greenhouses)
- Take clones from “mature” plants

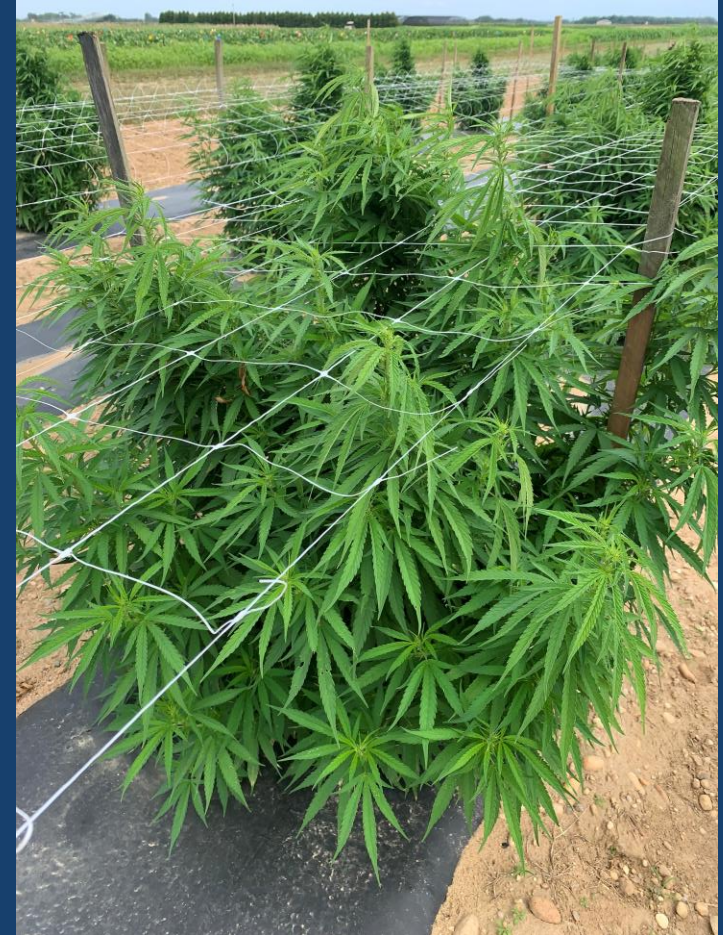


Hormodin #1 = 0.1% IBA (1,000 ppm)  
Hormodin #2 = 0.3% IBA (3,000 ppm)  
Hormodin #3 = 0.8% IBA (8,000 ppm)



# Stem Cutting Propagation of Hemp

- Take cuttings\* from a well-established and healthy plant (preferably not flowering)



*\*Don't forget to label your cuttings*



# Nutrition is important

- Stock plants fertigated with 100, 200, & 300 PPM N (15-5-15)
- Greatest fresh root weight and rooting percentages when cuttings are provided with nutrients in propagation
- Stock plants treated with 200 & 300 PPM N showed no chlorosis and produced larger plants (more harvestable cuttings)



From left to right: Stock plants treated with 100, 200, & 300 PPM N (15-5-15)



Alex Carver & Dr. James Faust  
Clemson University, South Carolina



# Stem Cutting Propagation of Hemp

- Remove apical cuttings from stock plant with 2 to 3 nodes



Approximately 5" – 8"

# Stem Cutting Propagation of Hemp

- Trim off any huge lower leaves and clip the top fan leaves if they are big



# Stem Cutting Propagation of Hemp

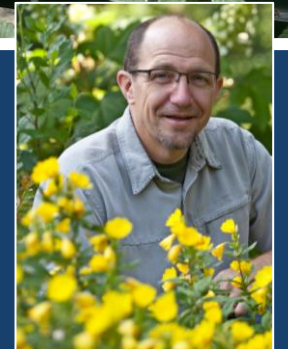
- Gently **wound** the bottom of the cutting
- Treat with a **rooting powder**
  - 0.1% IBA (1,000 ppm)





# Exogeneous Auxin Application

- Results are cultivar specific
  - 7 cv evaluated
- 4 x 4 factorial (16 treatment combinations) of K-IBA and K-NAA **liquid dip** treatments
  - 0, 2000, 4000, 6000 PPM K-IBA
  - 0, 2000, 4000, 6000 PPM K-NAA
- Overall, **2000-4000 ppm** NAA or IBA produced greatest rooting percentages when applied as quick dip



Alex Carver & Dr. James Faust  
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# Stem Cutting Propagation of Hemp

- Insert into a rooting medium



Campbell et al. (2021) report that **rockwool** offered a 7–13-fold improvement

# Stem Cutting Propagation of Hemp

- Place cuttings in a high humidity environment
  - Cloning domes
  - Intermittent mist





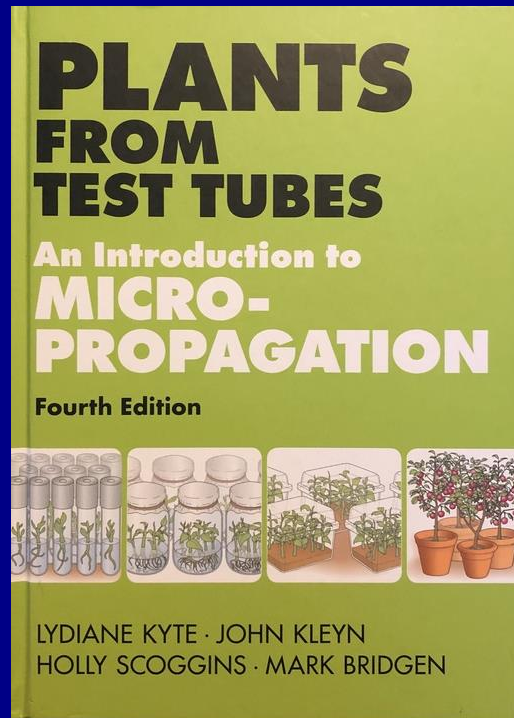
# Stem Cutting Propagation of Hemp

- About 72°-75°F are generally used. Plus 2°-3°F higher root zone temperature
- Long days: 16/8 (Light/Dark)



# Plant Tissue Culture

The growth of plants on a sterile nutrient medium *in vitro* under controlled environmental conditions



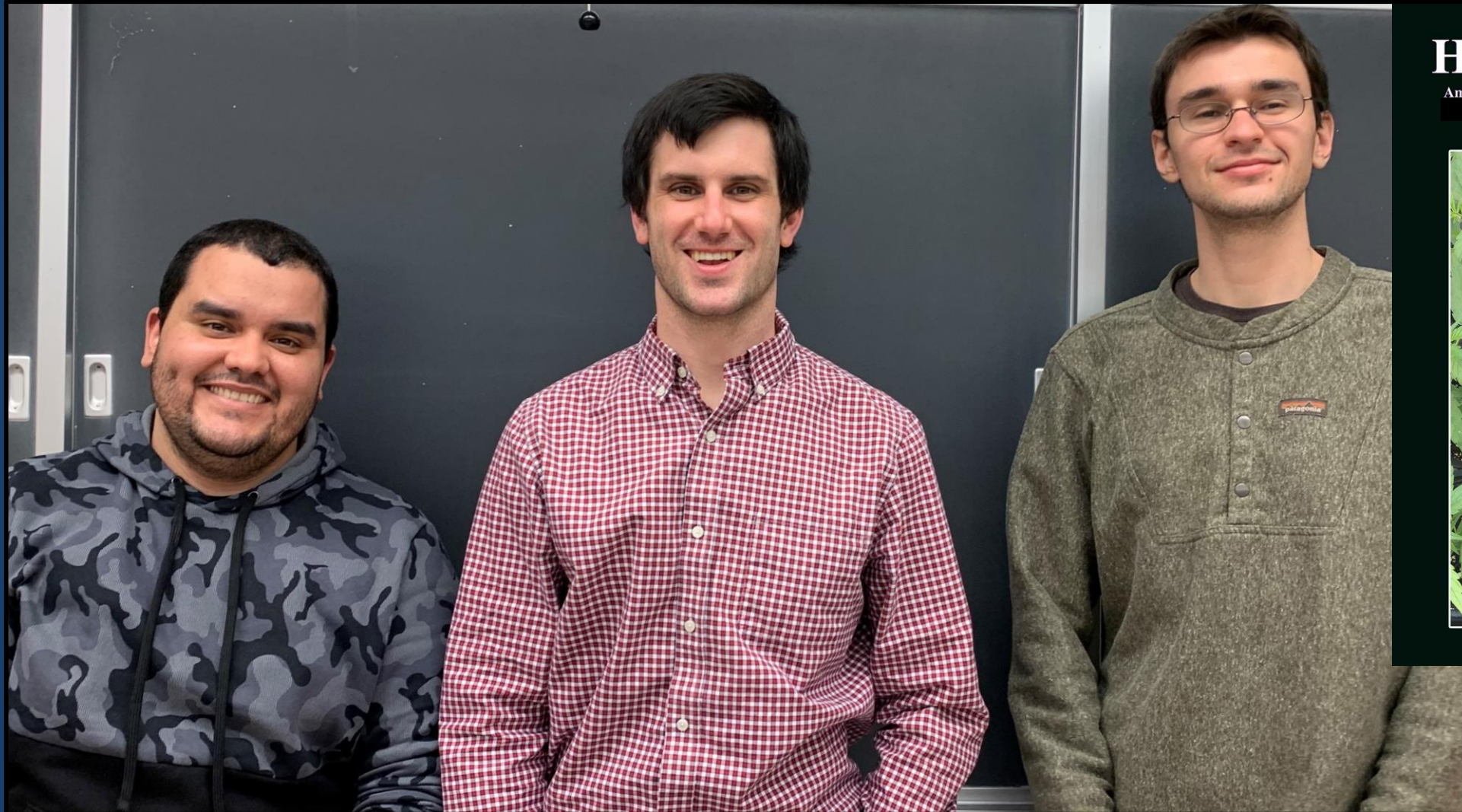
**MICROPROPAGATION**  
The rapid clonal propagation of plants *in vitro*





# Micropropagation of Hemp (*Cannabis sativa* L.)

## Victor Zayas, Conor Stephen, Andrei Galic



**HortScience**

American Society for Horticultural Science





# Why Hemp micropropagation?



- Clonal propagation
- Clean, vigorous plant material
- Efficient space utilization
- Large numbers
- Less maintenance
- Year-round production
- Controlled environment
- Disease free plants are possible

# Stages of Micropropagation



**Stage 0:**  
Proper Growth  
and  
Selection



**Stage I:**  
Introduction  
and  
aseptic  
establishment



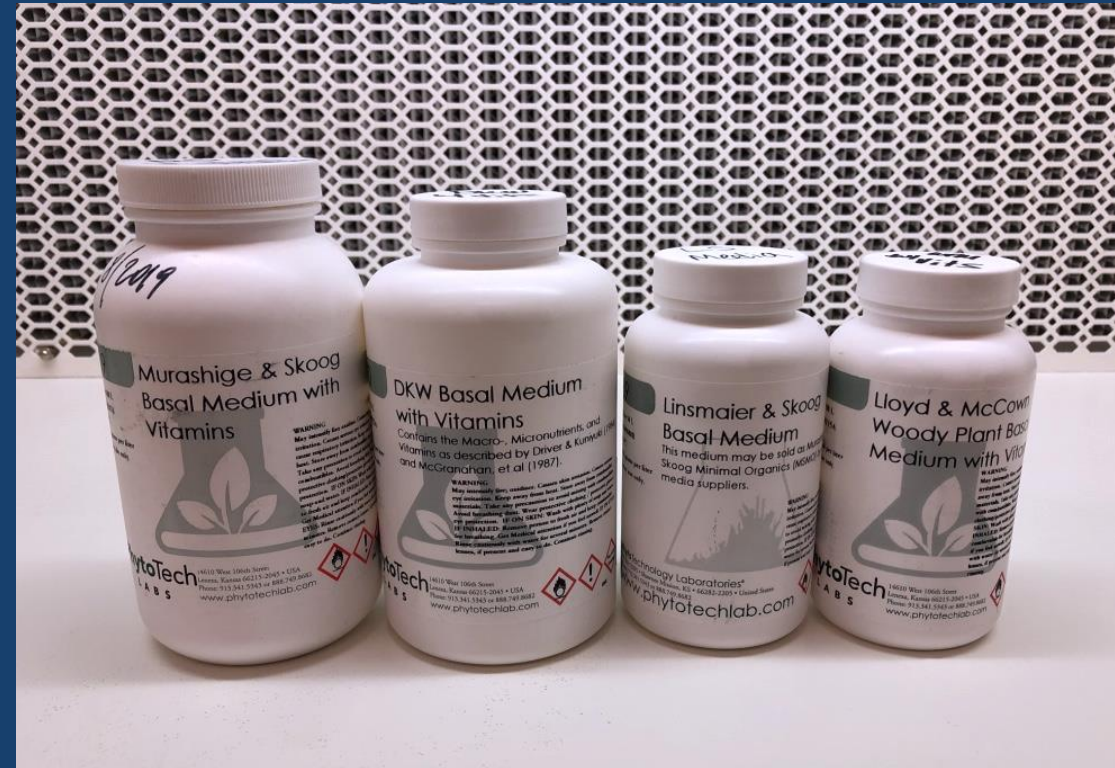
**Stage II:**  
Multiplication



**Stage III & IV:**  
Rooting/Acclimation

# Factors Influencing Micropropagation: Growth Medium

- Nutrients + vitamins
- Carbohydrates (Sugar)
- Gelling Agents
- Plant Growth Regulators (Hormones)





# Factors Influencing Micropropagation



- Explant type
- Temperature
- Culture pH

# Stage I Materials and Methods



'TJ's CBD'



1 cm shoot tips

## Treatments:

- Bleach concentrations (7.5% NaOCl)
  - 20%, 40%, 60%
  - 10 minute duration
- Stock plant environment
  - Greenhouse vs. Growth Chamber



# Stage I

Environment	Bleach Concentration	Percent Alive	Percent Contaminated
Greenhouse	20%	75%	90%
	40%	75%	90%
	60%	70%	85%
Growth Chamber	20%	100%	0%
	40%	95%	5%
	60%	100%	0%



## KEY POINTS:

- Plant material that is grown in a growth chamber has a better chance for success
- No differences observed between disinfection treatments – all work well
- No bleach concentration damaged plant material – suggesting higher concentrations can be used



# Stage II

## Nutrient Media



MS

LS

DKW

WPM

Murashige and Skoog (MS)  
Linsmaier & Skoog (LS)  
Driver & Kuniyuki Walnut Media  
(DKW)  
Lloyd & McCown Woody Plant  
Medium (WPM)

### Results:

- No significant differences between **MS, LS, and DKW** fresh weight, height, or rating
- **WPM** resulted in lowest fresh weight, shoot number, and rating

# Stage II

## Cytokinins

Comparing the cytokinins:

6-Benzylaminopurine (BA)

6-( $\gamma,\gamma$ -Dimethylallylamino) Purine (2iP)

Thidiazuron (TDZ)

at concentrations of

1.0  $\mu\text{M}$

5.0  $\mu\text{M}$

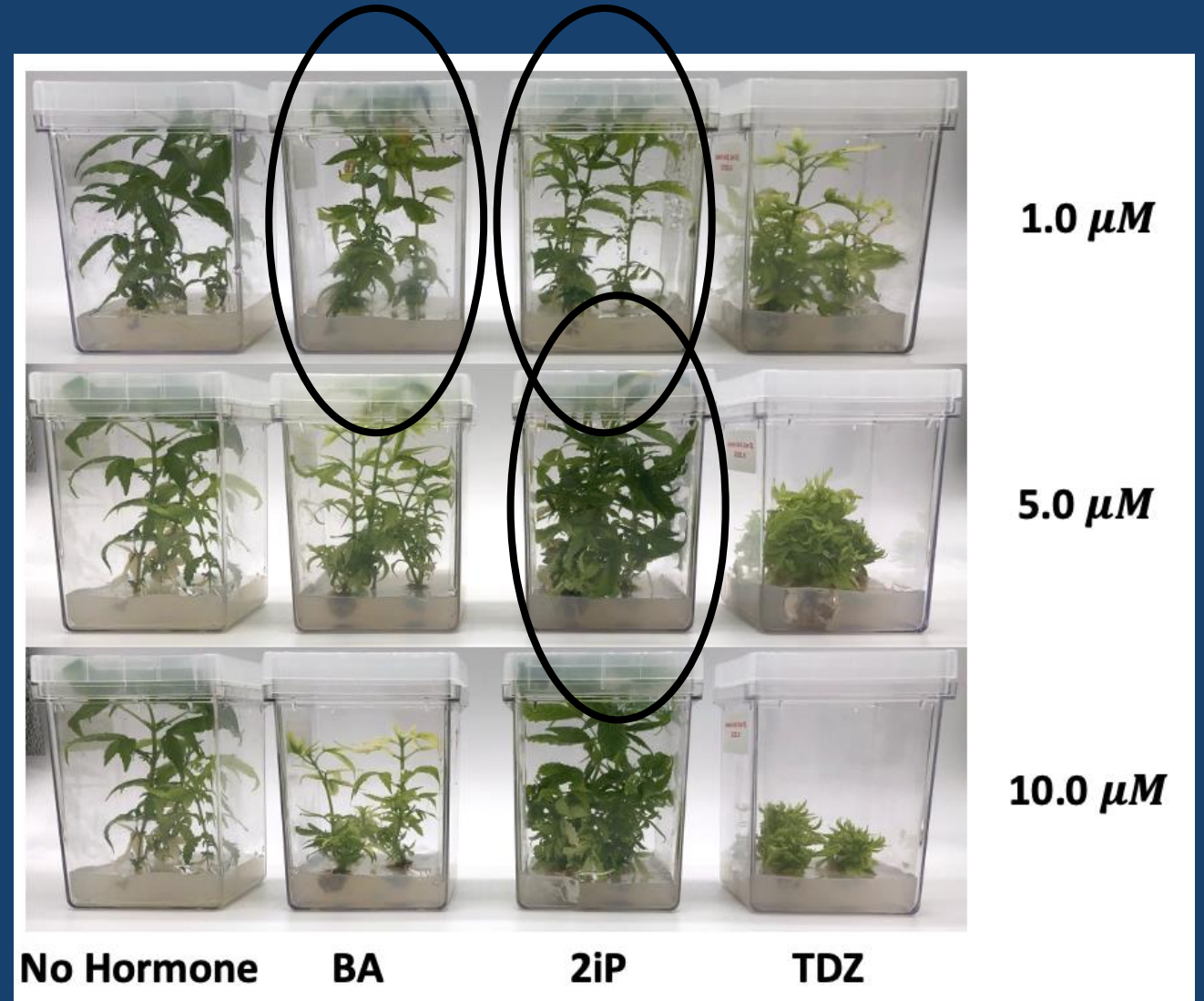
10.0  $\mu\text{M}$

# Stage II

## Cytokinin

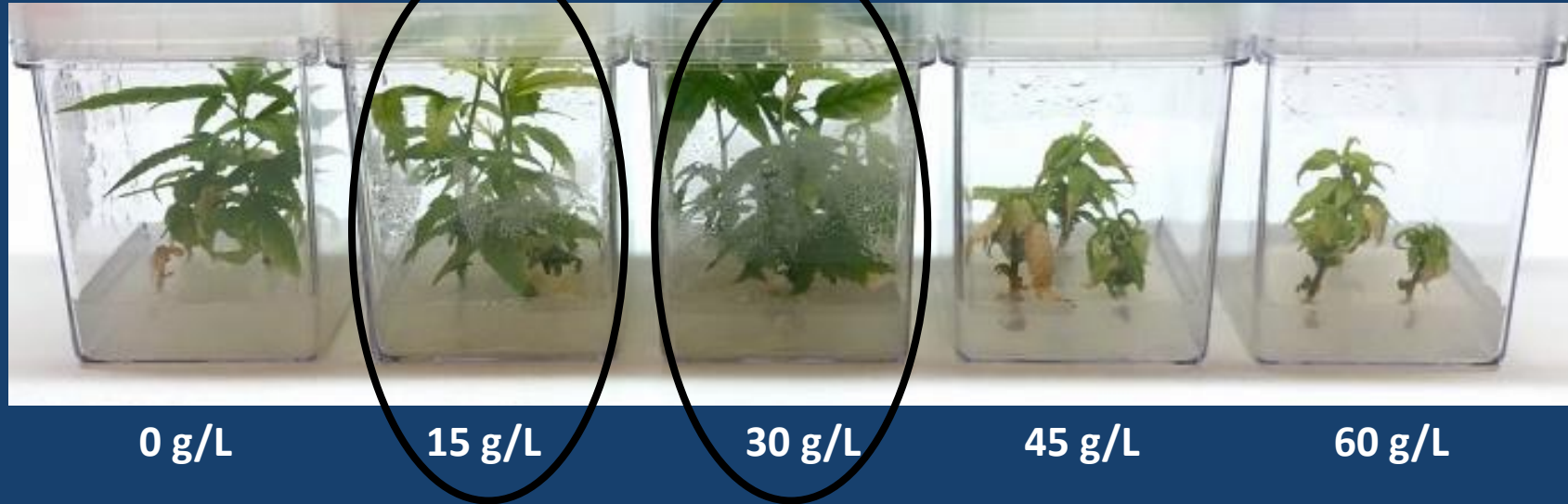
### Results:

- 5.0  $\mu\text{M}$  TDZ produced greater fresh weight and number of nodes
  - Also produced shorter shoots and more callus
- 1.0  $\mu\text{M}$  BA, 1.0  $\mu\text{M}$  2iP, and 5.0  $\mu\text{M}$  2iP produced significantly greater explant rating
- Low rates of 2iP or BA is recommended





# Stage II: Carbohydrate



## Results:

- 15 g/L and 30 g/L sucrose produced greater length, fresh weight, and rating
- 15 g/L showed less hyperhydricity
- 15 g/L sucrose recommended to maintain quality and reduce costs

# Stage II

## Gelling Agent



Agar

Agargellan

Gellan Gum\*

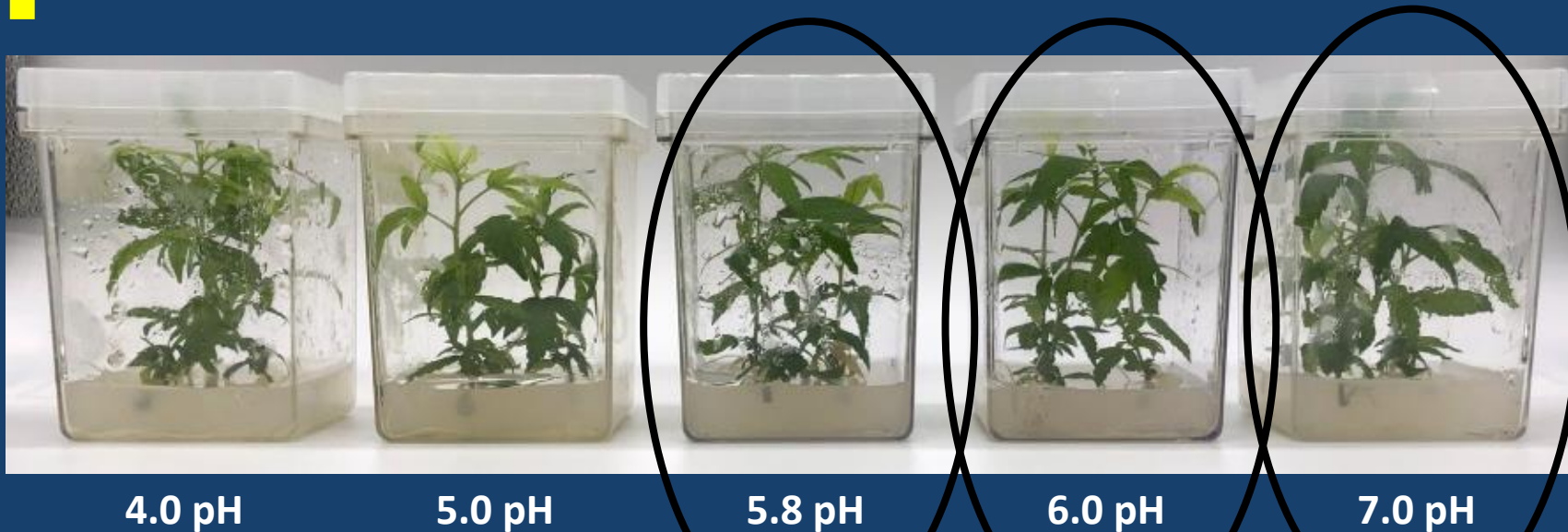
- **Results:**

- No differences in fresh weight, numbers of nodes, shoot length, and rating between treatments
- Long term exposure to gellan gum may be detrimental to growth

\* Gelrite, Phytigel, Gel-Gro, etc.

# Stage II

## pH



### Results:

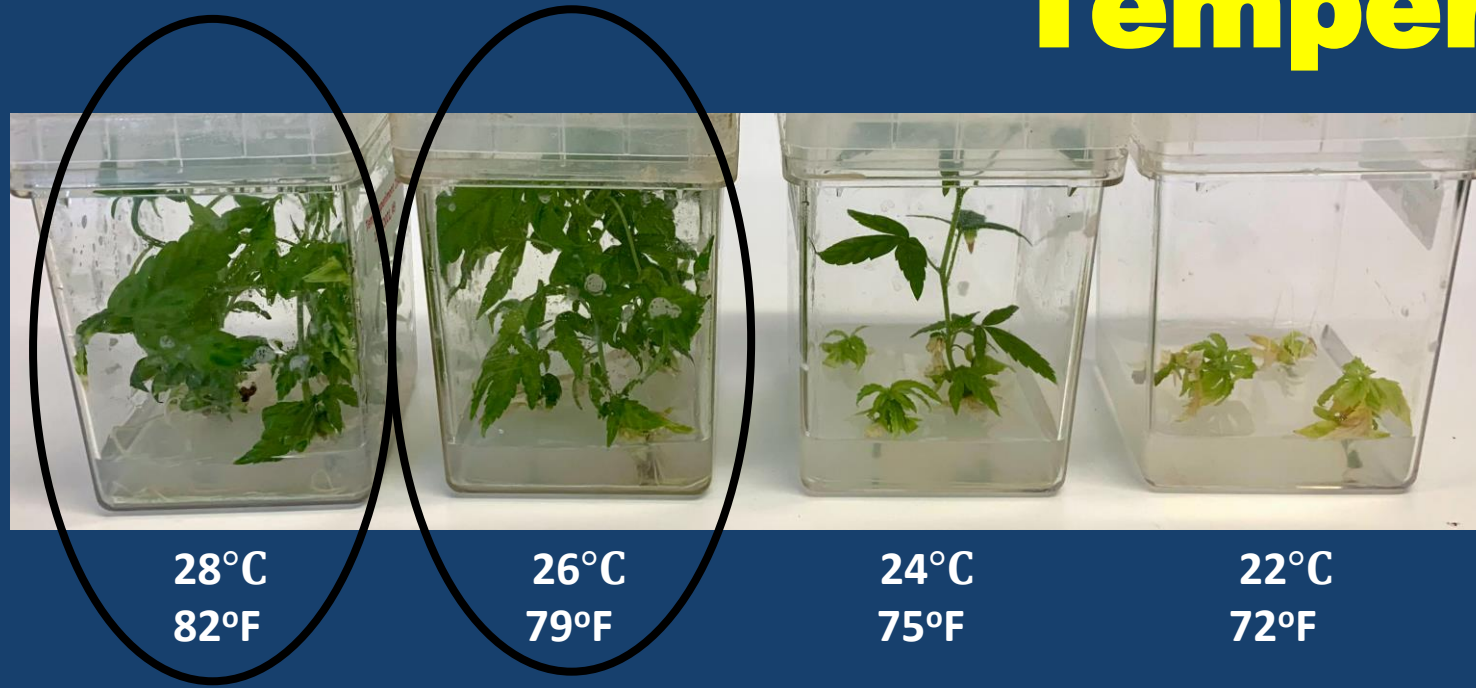
- 5.8, 6.0, and 7.0 pH produced more nodes and less chlorotic than 4.0 and 5.0 pH
  - No differences between treatment fresh weight, shoot length, and the number of explants that rooted
- The standard pH of 5.8 is recommended





# Stage II

## Temperature



### Results:

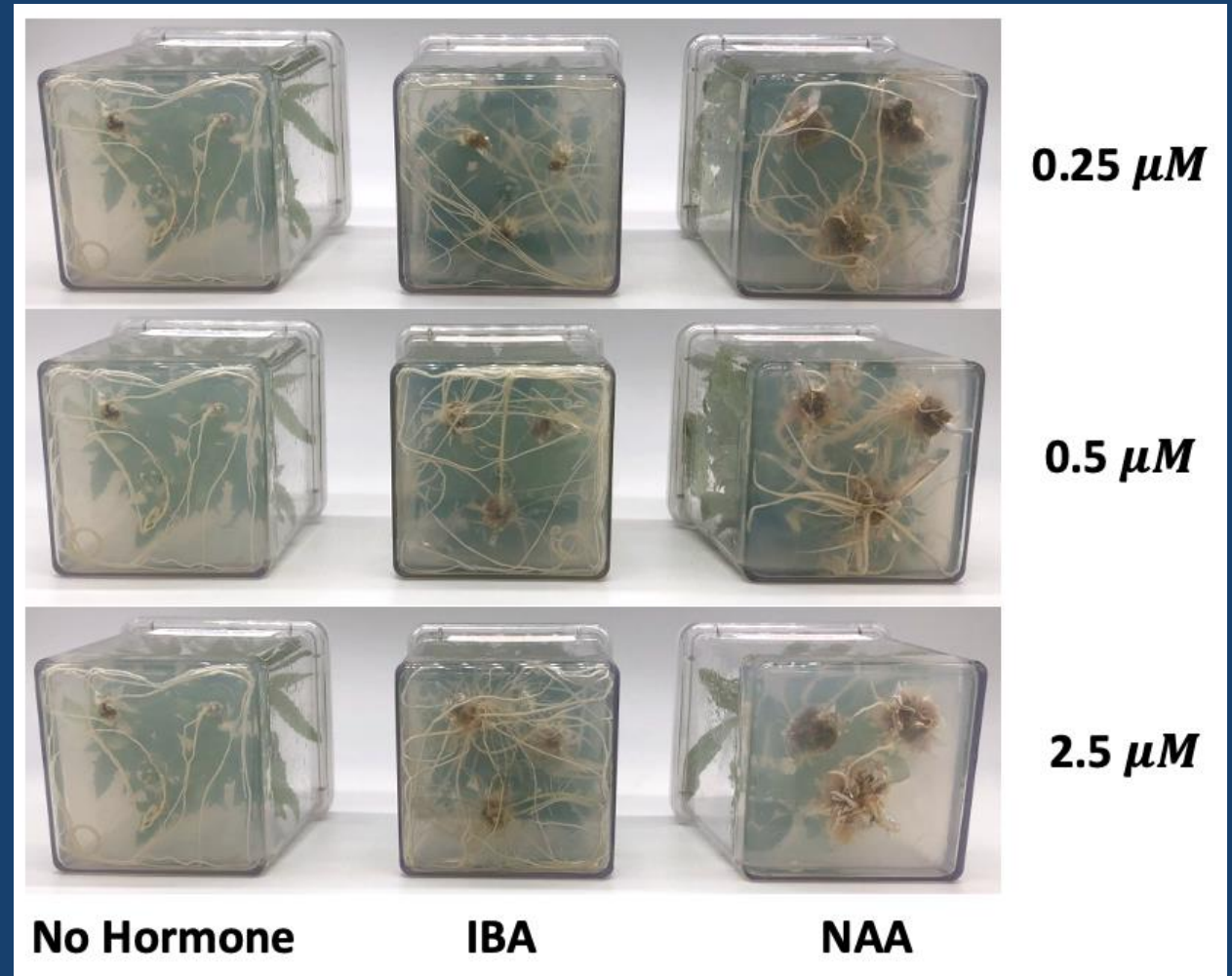
- 26°C and 28°C produced significantly greater nodes, roots, fresh weight, and rating
- Higher temperatures recommended for hemp micropropagation

# Stage III

## Auxin

### Results:

- 0.25  $\mu\text{M}$  NAA, 0.5  $\mu\text{M}$  NAA, and 2.5  $\mu\text{M}$  IBA produced greater:
  - shoot fresh weight
  - root fresh weight
  - number of nodes
- NAA produced more callus
- 2.5  $\mu\text{M}$  IBA recommended to stimulate root growth while maintaining shoot quality *in vitro*



# Stage IV Acclimation

Dome vs. intermittent mist  
Auxin vs. no auxin

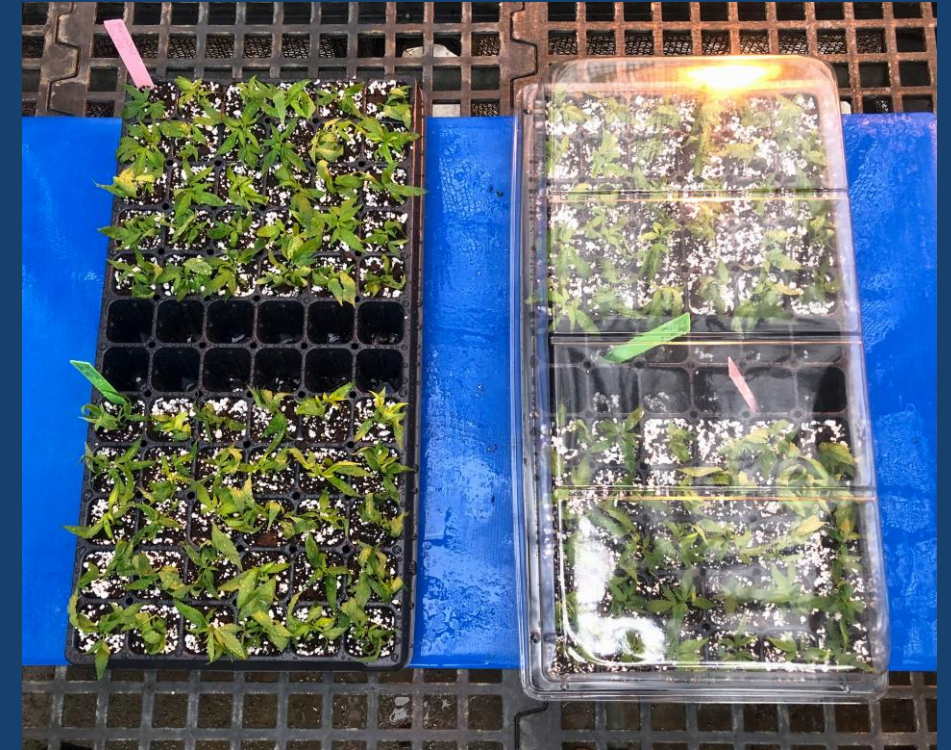
Plastic Domes



Auxin



Intermittent Mist





# Stage III & IV Acclimation

Auxin

No Auxin



No Dome

Dome



# Conclusions

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- Cytokinin type and concentration affect *in vitro* growth
- Stage III is not necessary
- Plants that are acclimated under plastic domes perform better than intermittent mist
- Auxin assists root production, but not necessary for success
- Gelling agent and pH appear to have the least impact on growth compared to other variables that were tested





# *Examining the stages of micropropagation for hemp*



**Conor Stephen**, MS 2022  
**Victor Zayas**, PhD candidate  
**Andrei Galic**, MPS 2021  
**Victoria Zeng**, BS 2022

# QUESTIONS?

